# Trophic Interactions Workgroup Update

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**SPIS-CRISPS-WO#5** Trophic Interactions



### **Trophic Interactions Workgroup Objectives**

- 1. Identify the major algal grazers and their consumers. \*Food Web Structure
- 2. Determine algal growth and grazing rates of small grazer species. \*Species Interactions
- Assess the potential for top-down (consumer) control of key grazers in the ecosystem.
   \*Consumer Effects

# **Objective 1: Food web Structure**

1. What are the major pathways of energy flow and material transport?

2. Which grazers consume benthic filamentous algae (a.k.a, nuisance algae) and to what degree?

3. Which predators consume algal grazers?

### **Objective 1: Methods**

### Stable Isotope Analysis-SIA (δ<sup>13</sup>C & δ<sup>15</sup>N)

- Integrated signal of consumers' dietary choices
- Isotopic mixing models provide estimates of the proportional dietary contributions from discrete resource pools

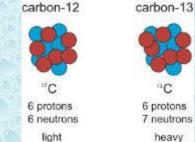
- Stomach/Scat Content Analysis-SCA
  - 'snap-shot' of diet in time
  - Confirm predator-prey links: Who's eating who?
  - Inform isotopic models: 'prior information'

## Stable Isotope Analysis-SIA

Ratio of heavy to light isotopes (<sup>13</sup>C:<sup>12</sup>C, <sup>15</sup>N:<sup>14</sup>N)

 $\delta X$  (‰) = [ $\mathbf{R}_{sample} \mathbf{R}_{standard} - 1$ ]×1000, where X is element of interest

You are what you eat (± discrimination,  $\Delta X_{tissue-diet}$ )



6 protons 7 neutrons heavy

#### $δ^{13}$ C has Small discrimination $Δ^{13}$ C<sub>tissue-diet</sub> ≈ 1.0‰ ± 0.5

-Differs among plants with different photosynthetic pathways (i.e., C3, C4, CAM, etc.)

-Varies in aquatic producers due to δ<sup>13</sup>C of dissolved inorganic carbon-DIC sources, δ<sup>13</sup>C- $CO_2aq$  and  $\delta^{13}C-HCO_3^{-1}$ , as well as relative concentrations of [CO<sub>2</sub>aq], and [HCO<sub>3</sub><sup>-1</sup>].

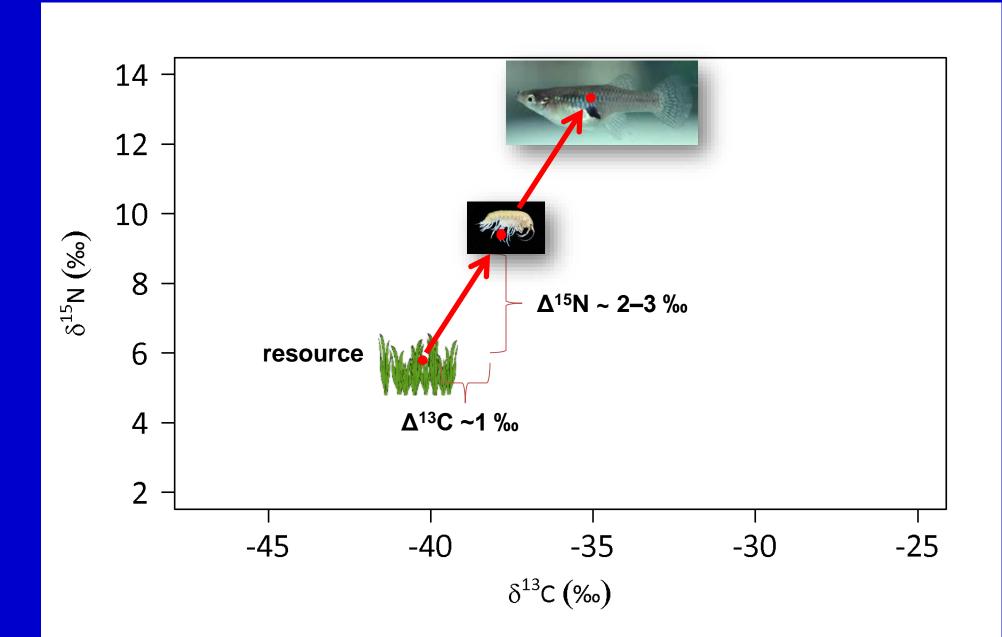
-Indicator of carbon pools (resource categories) used by consumers

 $\delta^{15}N$  has Larger discrimination  $\Delta^{15}N_{tissue-diet} \approx 2.2 \% \pm 0.7$ 

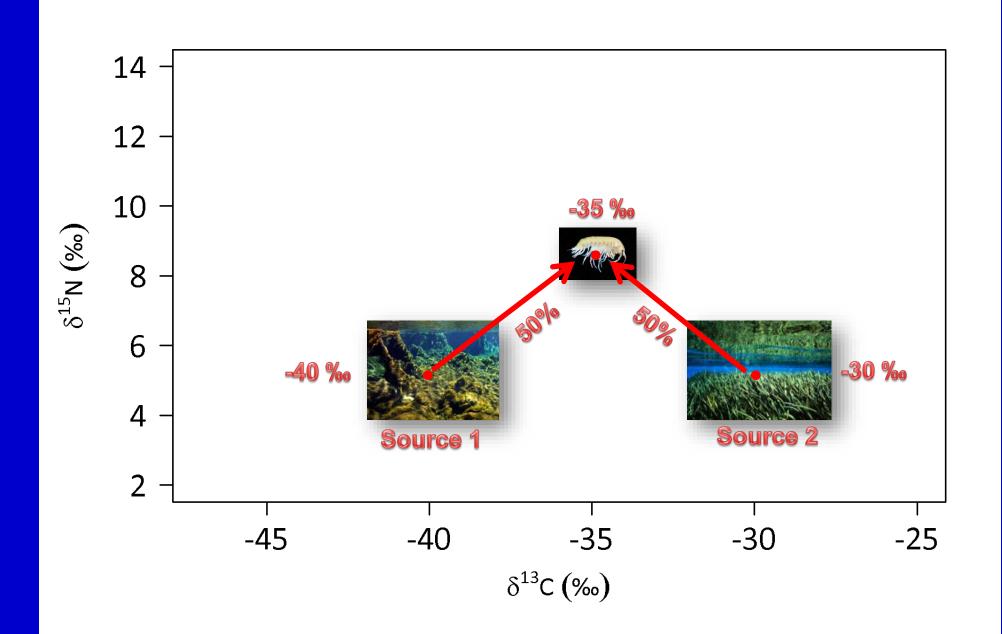
-Indicator of nitrogen sources and cycling processes at food web base

-Quantify trophic level of consumers

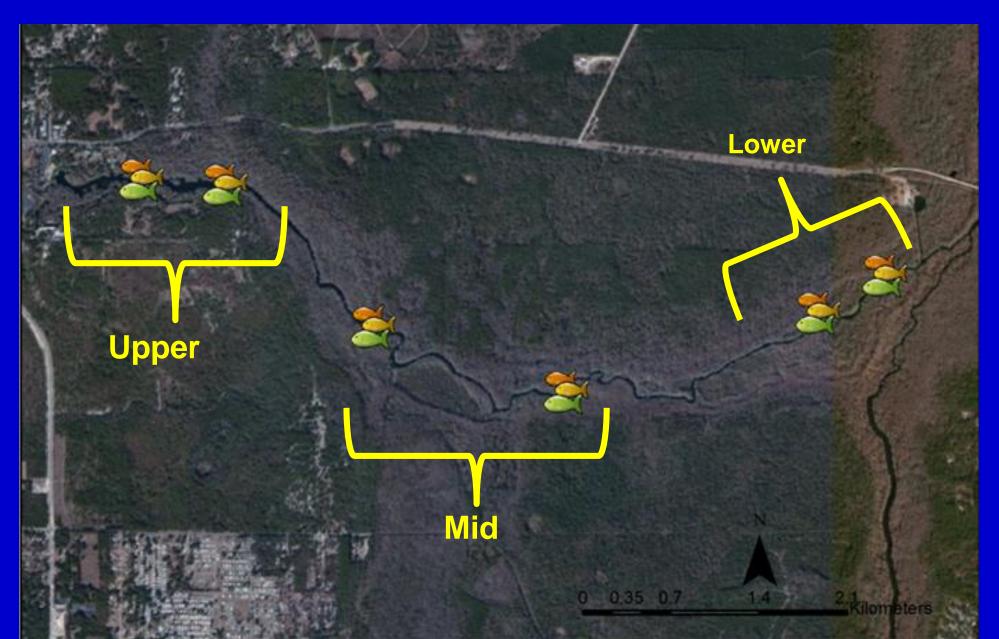
### **Isotopic Discrimination:** ΔX<sub>tissue-diet</sub>



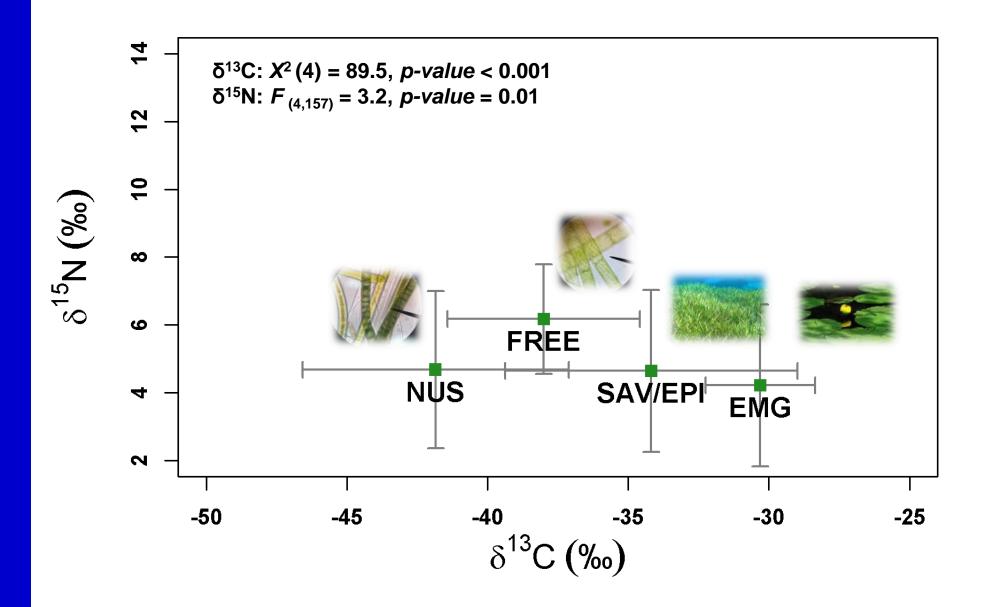
#### **Isotopic Mixing: Multiple Sources**



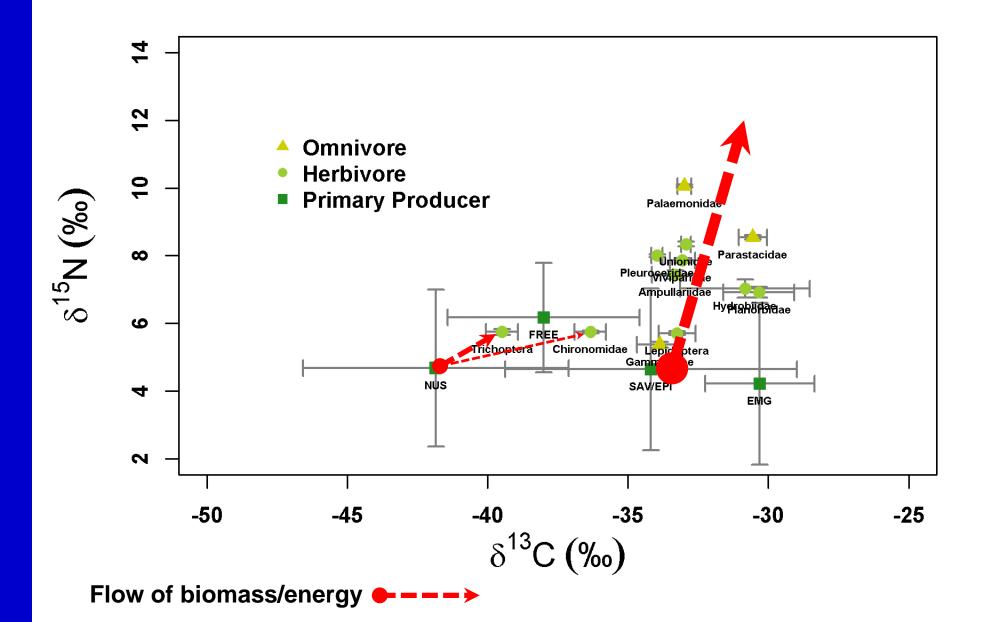
# **Sampling Sites**



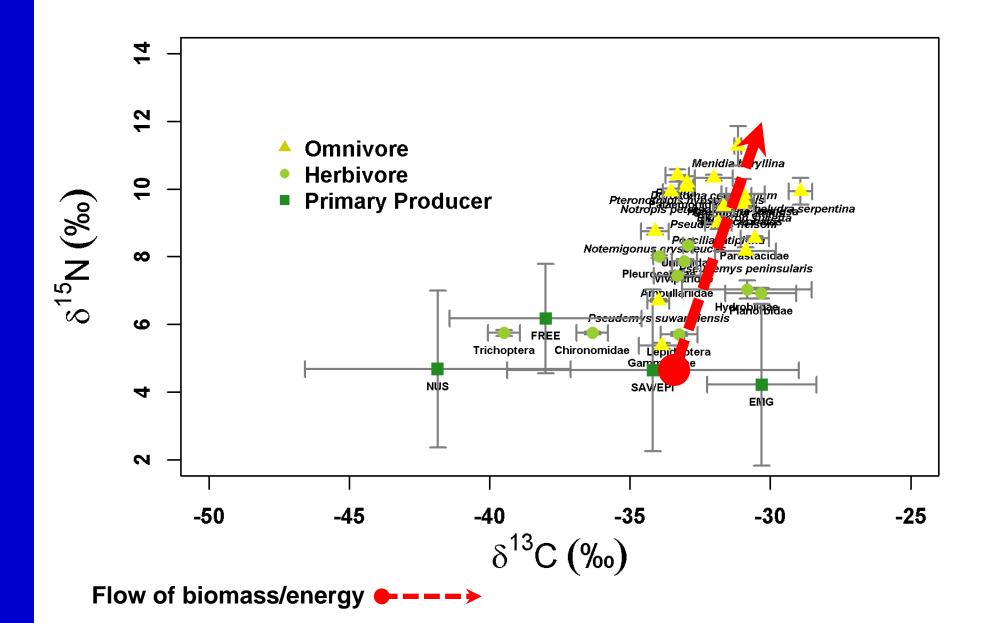
#### **Primary Producer Groups (Sources)**



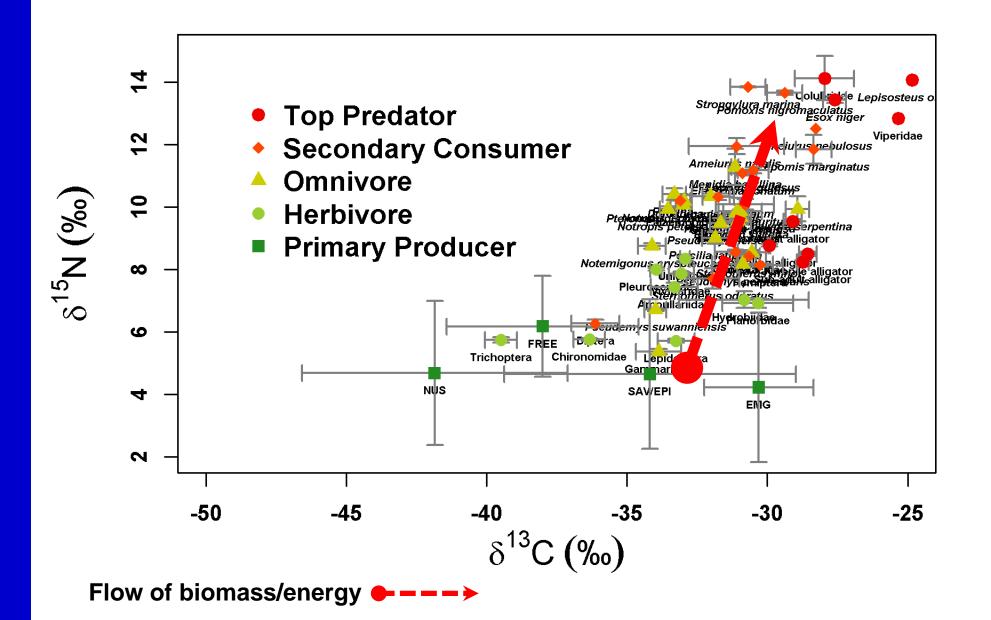
### Herbivores and Omnivores (inverts)



### **Herbivores and Omnivores**

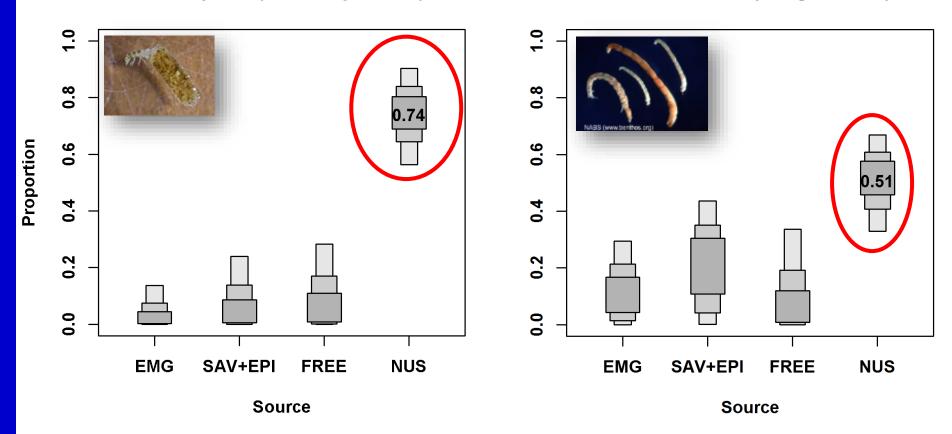


### **Secondary Consumers and Top Predators**



# **Isotopic Mixing Model Results**

Trichoptera (caddisfly larvae)



Chironomidae (midge larvae)

What about predators? Who is eating the algal grazers?

# Stomach Content Analysis (SCA) Fish













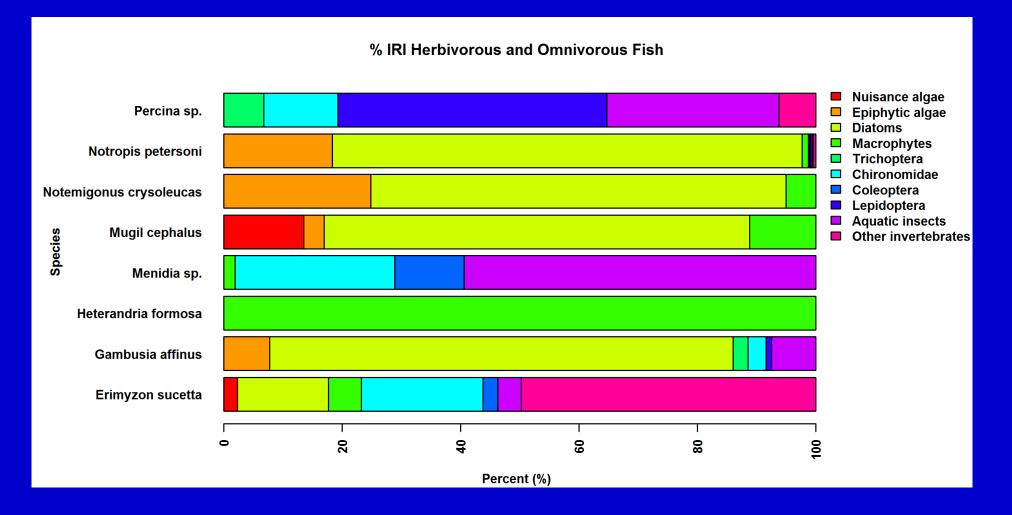








Turtles (scat)

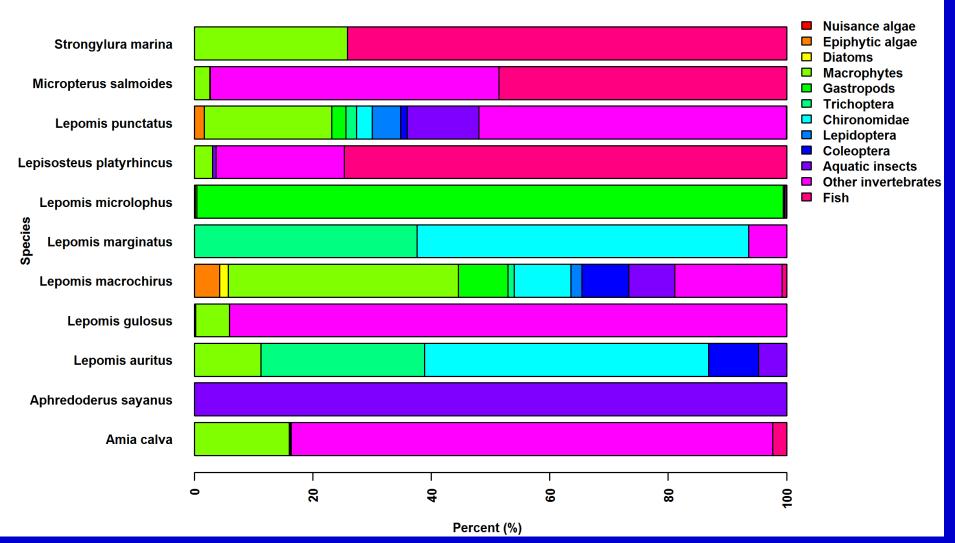


1. Diatoms highly important resource.

2. Trichopterans and chironomids relatively unimportant prey.

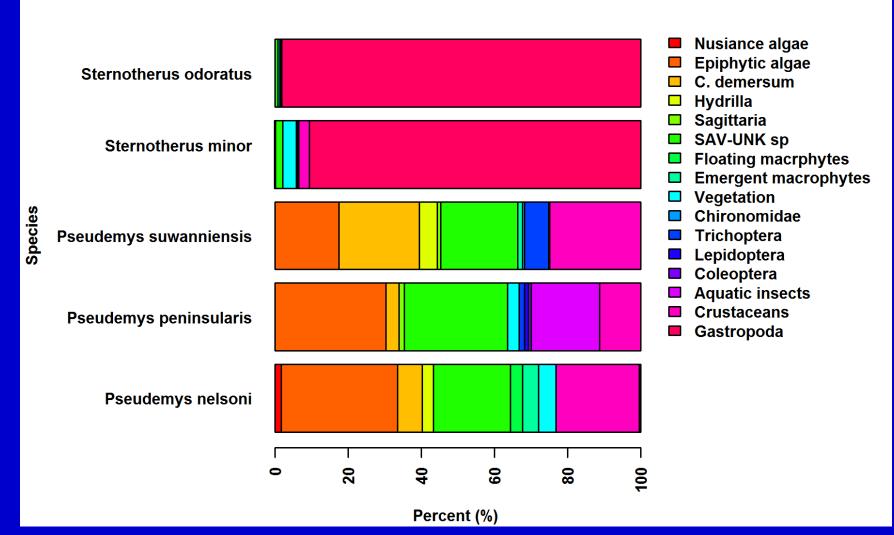
3. Little evidence of nuisance algal consumption.

#### % IRI Predatory Fish



- 1. L. marginatus and L. auritus major predators of trichopterans.
- 2. L. microlophus major gastropod predator.
- 3. Other invertebrates (i.e., decapods , amphipods) and fish are primary prey for most species

#### % IRI Turtles



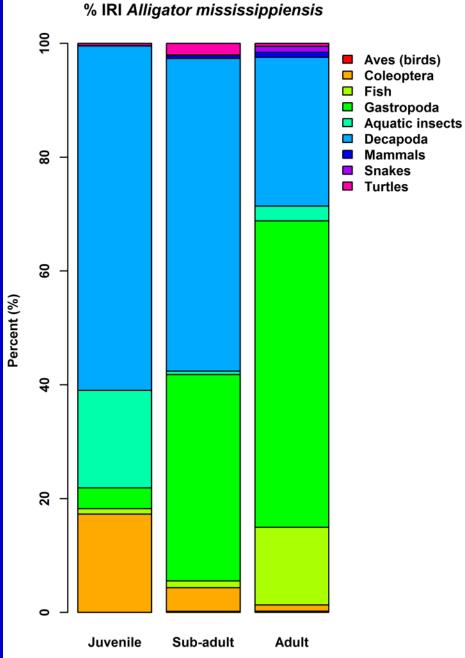
- 1. S. odoratus and S. minor chiefly predators of small benthic gastropods (i.e., physids, hydrobiids, planorbids)
- 2. River cooters (*Pseudemys* spp.) mainly consume macrophytes and to lesser extent small invertebrates.

# Alligators









# **Objective 1: Conclusions**

- Nuisance filamentous contributes little to aquatic food web
- Few grazers heavily rely of nuisance algae
  Invertebrates:

Trichopterans>Chironomids>Rhagionids>Amphipods>Lepidopterans>Gastropods

- Vertebrates:

**Shiners > Darters** 

- Major predators of algal grazers include Redear Sunfish, other Sunfish species, and kinosternid turtles
- Alligators are not 'Apex predators' rather they primarily feed on species occupying lower trophic levels (i.e., gastropods, decapods, insects)

# **Objective 2: Species Interactions-Grazing**

1) Will gastropod and decapod grazers consume filamentous algae?

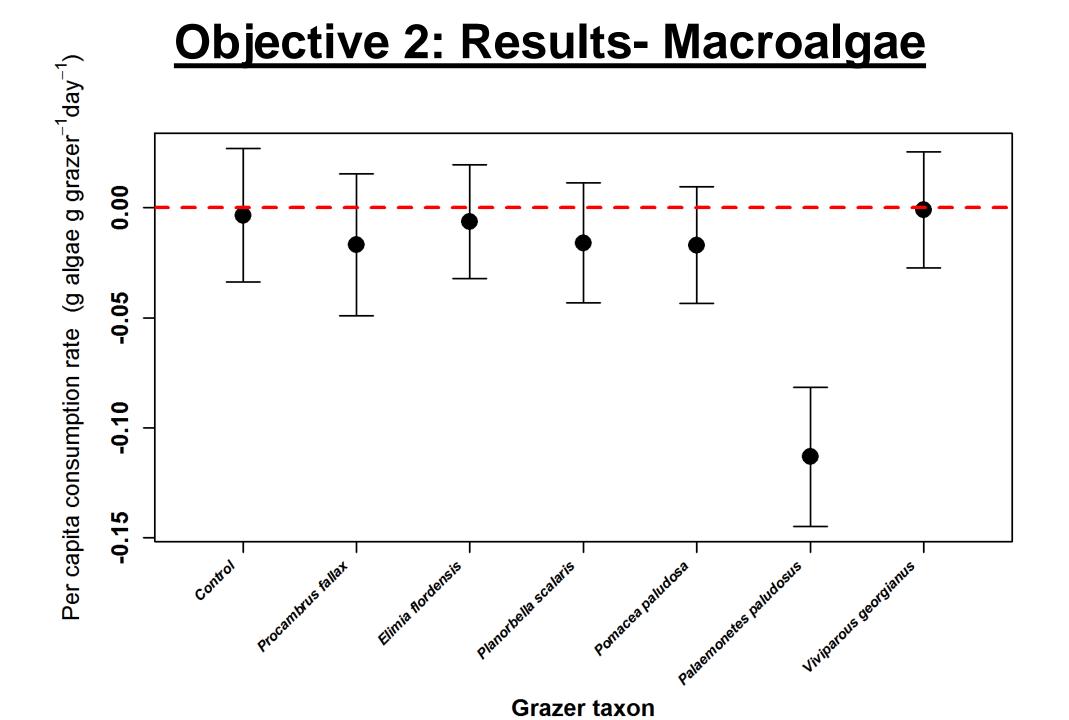
### 2) Is there potential for these grazer species to exert topdown control over filamentous algae?

- Grazer taxa: Elimia floridensis, Viviparus georgianus, Pomacea paludosa, Planorbella scalaris, Palaemonetes paludosus, and Procambarus fallax
- Macroalgae: Lyngbya, Vaucheria, Spirogyra, Rhizoclonium, Cladophora, and mixed Rhizoclonium + Cladophora
- Submerged macrophytes (SAV): Hydrilla verticillata, Ceratophyllum demersum, Sagittaria kurziana, Vallisneria americana, and Najas guadalupensis.

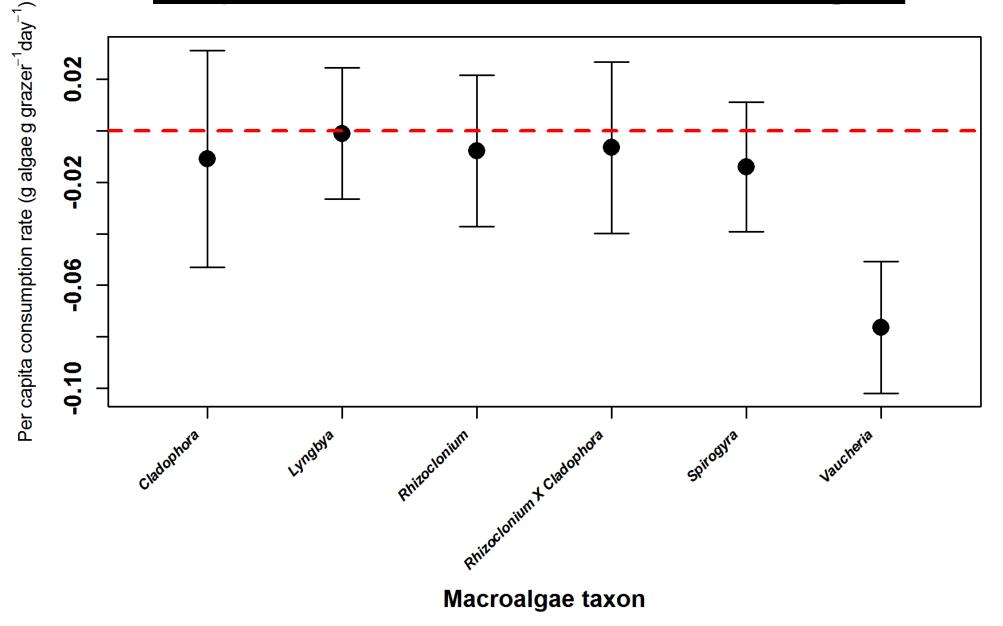
# **Objective 2: Grazer Experiments**



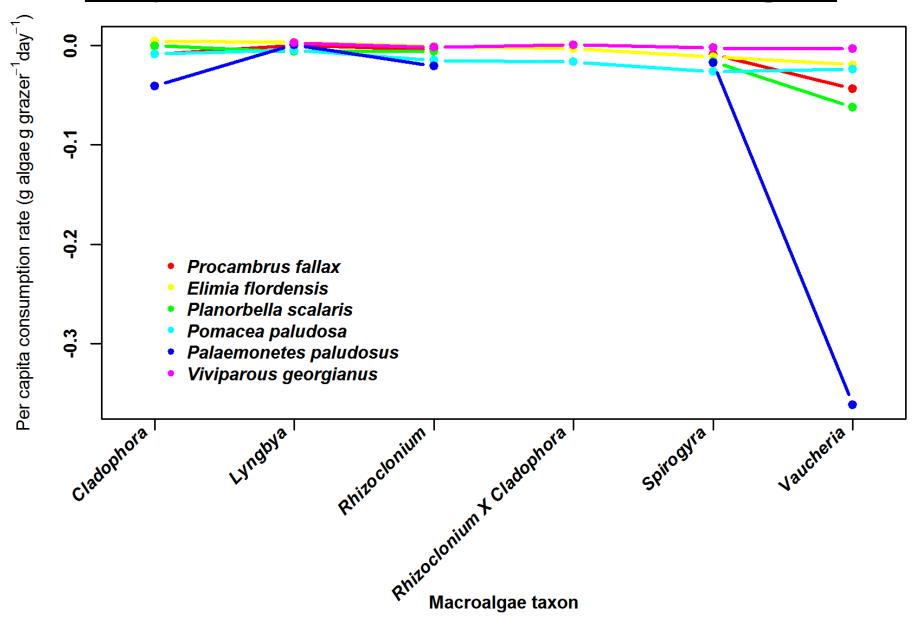




### **Objective 2: Results- Macroalgae**



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# **Objective 2: Summary-Macroalgae**

• Grazer capacity =

Grass shrimp > Apple snail > Crayfish > Planorbids > Other gastropods



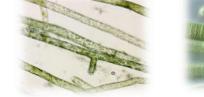
• Macroalgae Preference =

Vaucheria > Spirogyra > Rhizoclonium X Cladophora > Rhizoclonium> Lyngbya > Cladophora



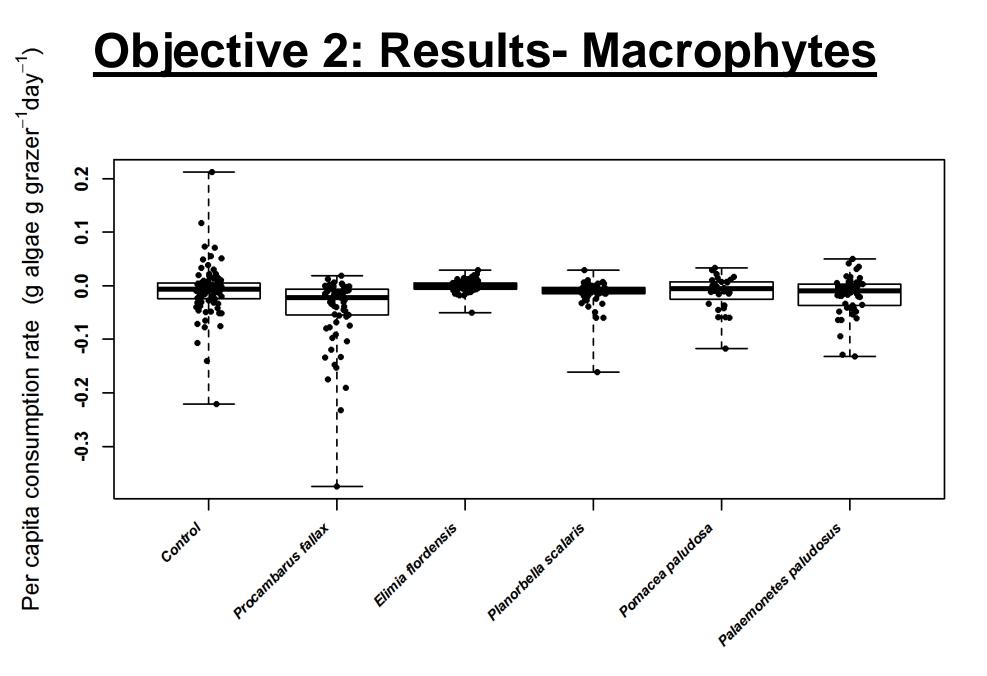




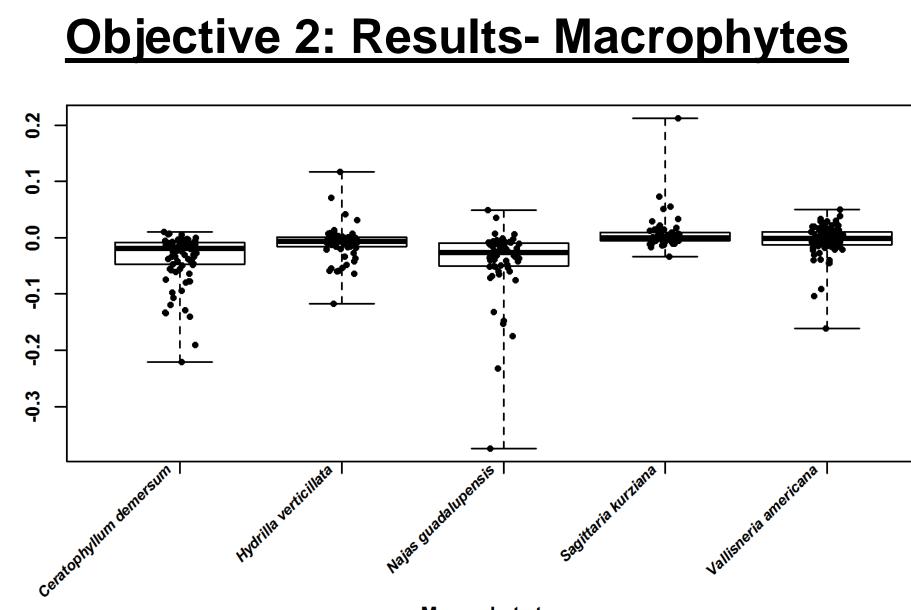








Grazer taxon



Macrophyte taxon

# **Objective 2: Summary-Macrophytes**

• Grazer capacity =

### Crayfish > Grass shrimp > Apple snail > other gastropods











Macrophyte preference =

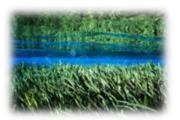
Najas > Ceratophyllum > Hydrilla > Sagittaria > Vallisneria









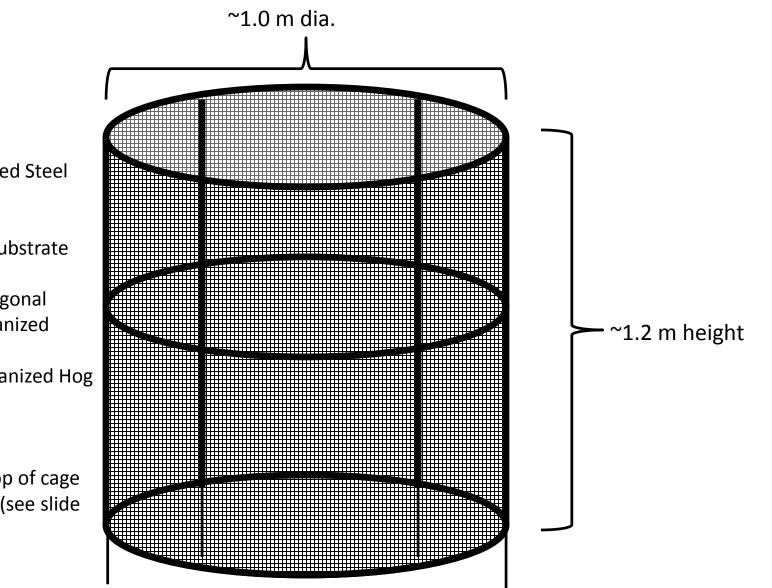


# **Objective 3: Consumer Effects**

- 1. What is the impact of removing predation pressure on the herbivore community, particularly aquatic insects, crustaceans, and gastropods?
- 2. Is more epiphytic algal biomass consumed by grazers in the absence of predation?

3. Does SAV benefit?

# **Exclusion Cage Design**



#### <u>Frame</u>

-0.625 cm Galvanized Steel Rods and hoops -All welded -rods extend into substrate <u>Mesh</u> -2.5 cm mesh Hexagonal -Vinyl Coated Galvanized Steel -Secured with Galvanized H

-Secured with Galvanized Hog Rings

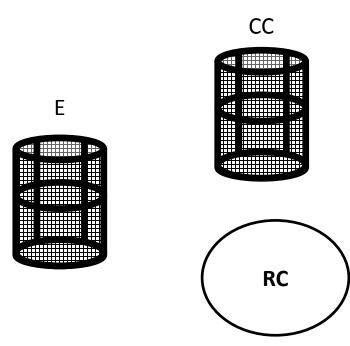
#### <u>Signage</u>

-Flagging tape at top of cage-Sign at each array (see slide4)



## Cage Array Design

### 2 cages per array

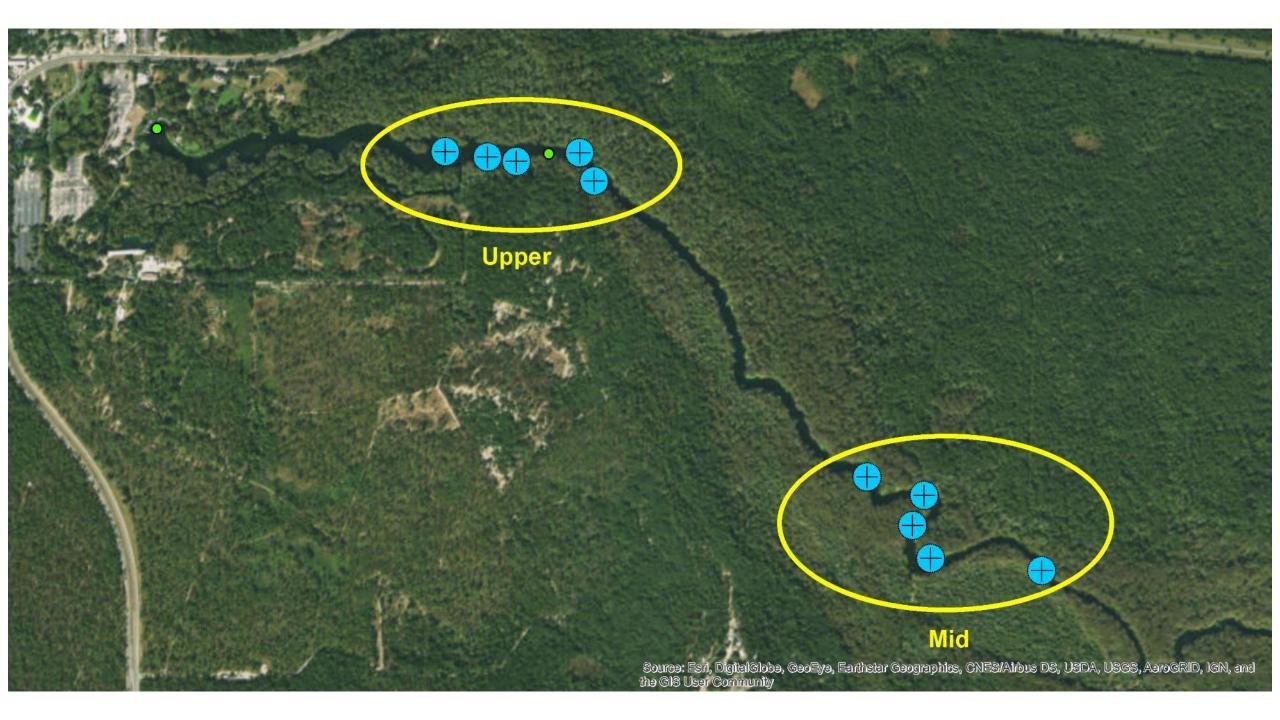


### <u>Array</u>

- -1 Exclusion Cages-E
- -1 Cage Control-CC
- -1 Reference Control-RC

\*Cage Control treatment will have entire panels of mesh removed from cage to allow organisms uninhibited access while replicating shading and flow effects of true exclusion cages (basically an exclusion cage with ½ the mesh removed).

\*Reference Control is simply a monitoring area with same spatial footprint as cages.









## Cages may be fully submerged



### Cages may protrude from water's surface



# <u>Schedule</u>

- Installed and initial sampling: Oct. 2016
  - Processing samples (SAV biomass, epiphytic algae, and invertebrate abundance and biomass).

• Monitoring once per week to clean/repair cages

• SAV growth: March/April 2017

• Breakdown and final sampling: April/May 2017

## Questions?