### From algae to alligators: Examining food web structure in Florida's spring ecosystems

#### James C. Nifong<sup>1</sup>, Tom Frazer<sup>2</sup>, and Rob Mattson<sup>3</sup>

<sup>1</sup>Fisheries and Aquatic Sciences, University of Florida, Gainesville, FL <sup>2</sup>School of Natural Resources and Environment, University of Florida, Gainesville, FL <sup>3</sup>St. Johns River Water Management District, Palatka, FL

Email: ncboy@ufl.edu

**CRISPS Work Group 5: Trophic Interactions** 







1. Research Questions

2. Methods

3. Results

4. Conclusions

5. Preliminary Data: Grazing Trials

6. Future Directions: Exclusion Experiments

# **Research Questions**

1. What are the major pathways of energy flow and material transport in Florida spring ecosystems?

2. Which grazers consume benthic filamentous algae (a.k.a, nuisance algae) and to what degree?

3. Which predators consume algal grazers?

### **Methods**

- Stable Isotope Analysis-SIA (δ<sup>13</sup>C & δ<sup>15</sup>N)
  - Integrated signal of consumers' dietary choices
  - Isotopic mixing models provide estimates of the proportional dietary contributions from discrete resource pools

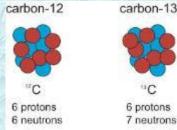
- Stomach/Scat Content Analysis-SCA
  - 'snap-shot' of diet in time
  - Confirm predator-prey links: Who's eating who?
  - Inform isotopic models: 'prior information'

## **Stable Isotope Analysis-SIA**

Ratio of heavy to light isotopes (<sup>13</sup>C:<sup>12</sup>C, <sup>15</sup>N:<sup>14</sup>N)

 $\delta X$  (‰) = [ $\mathbf{R}_{sample} / \mathbf{R}_{standard} - 1$ ]×1000, where X is element of interest

You are what you eat (± trophic enrichment,  $\Delta X_{tissue-diet}$ )



heavy

light

 $\delta^{13}$ C has small trophic enrichment  $\Delta^{13}$ C<sub>tissue-diet</sub>  $\approx 1.0\% \pm 0.5$ 

-Differs among plants with different photosynthetic pathways (i.e., C3, C4, CAM, etc.)

-Varies in aquatic producers due to  $\delta^{13}C$  of dissolved inorganic carbon-DIC sources,  $\delta^{13}C$ - $CO_2aq$  and  $\delta^{13}C$ - $HCO_3^-$ , as well as relative concentrations of [ $CO_2aq$ ], and [ $HCO_3^-$ ].

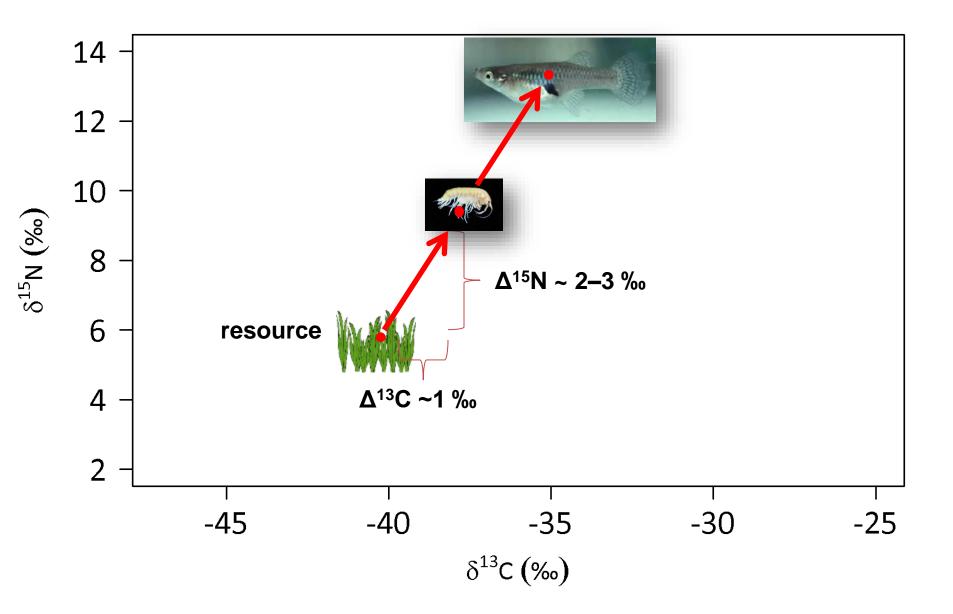
-Indicator of carbon pools (resource categories) used by consumers

 $\delta^{15}$ N has larger trophic enrichment  $\Delta^{15}$ N<sub>tissue-diet</sub>  $\approx 2.2 \% \pm 0.7$ 

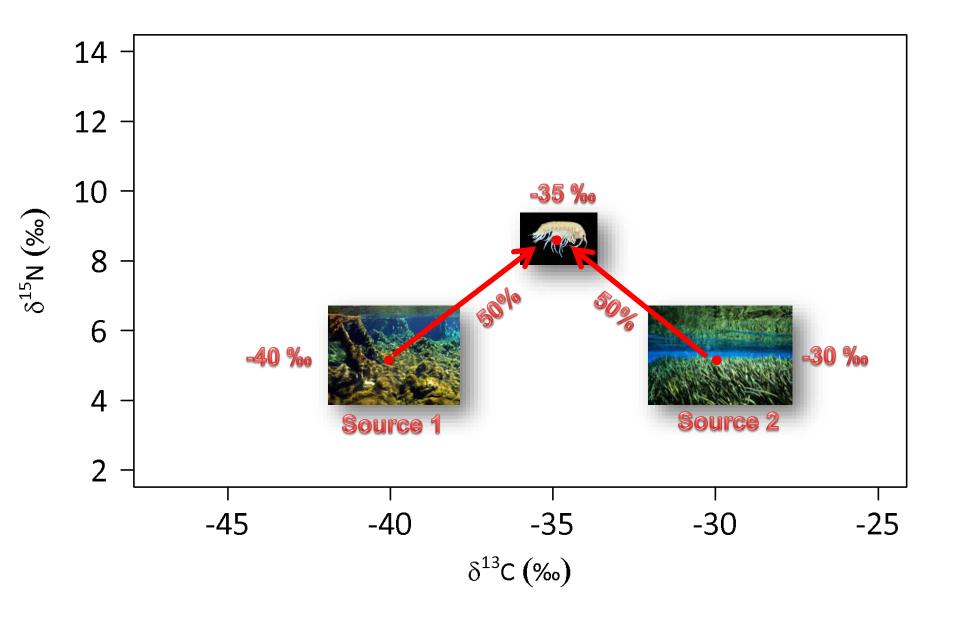
-Indicator of nitrogen sources and cycling processes at food web base

-Quantify trophic level of consumers

### **Trophic Enrichment: ΔX**<sub>tissue-diet</sub>



### **Isotopic Mixing: Multiple Sources**



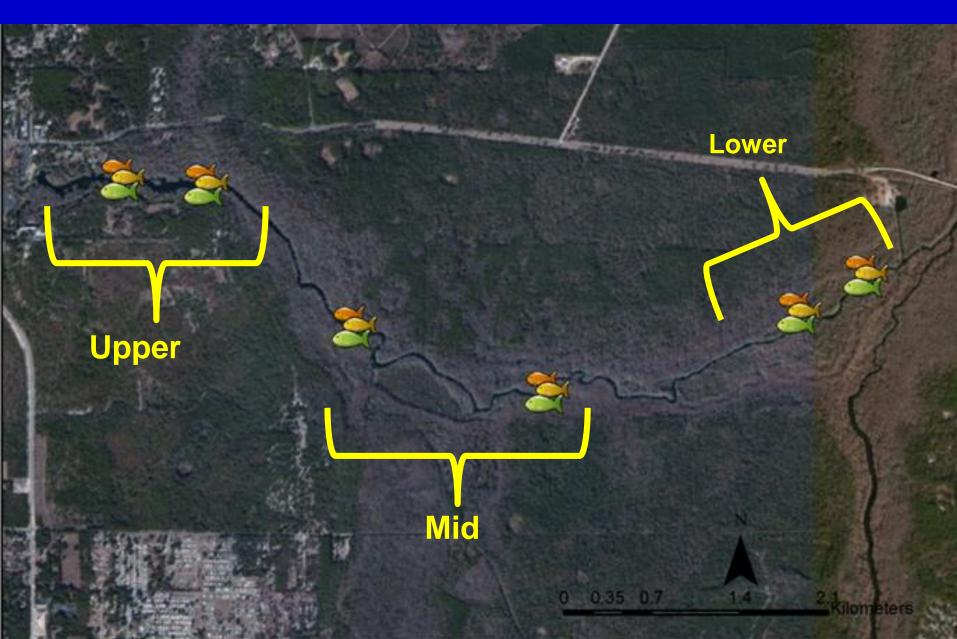
### Silver Springs/Silver River



**Mammoth Spring** 

Silver River State Park Ocklawah a River

# **Sampling Sites**



# **Primary Producers**

### <u>Submersed Aquatic Vegetation</u> (SAV) Sagittaria kurziana

Vallisneria americana

Hydrilla verticillata

#### Ceratophyllum demersum

#### Macroalgae and diatoms

Benthic Filamentous Algaea.k.a Nuisance Algae

Lyngbya

Vaucheria

**Unattached Algae** 

Spirogyra

Epiphytic Filamentous Algae and Diatoms



#### Emergent and Floating Vegetation



Nuphar avenda (spatterdock)

> Sagittaria lancifolia (arrowhead)





### **Herbivores and Omnivores (Inverts)**

Trichoptera

(caddisfly)

### **Emergent Insects:**

### Gastropods:

**Hydrobiids** 

(mud snail)



Physids (silt snail)



Pleurocerids (elimia snails)



Planorbids

(ramshorn snail) Vivip ) (myste

NABS (www.benthos.pro

Chironomidae (midge)



<sup>II)</sup> Viviparids (mystery snail)



Ampullariids (apple snail)

### **Crustaceans:**



Gammaridae (amphipod)



Palaemonids (grass shrimp)



Lepidoptera

(moth)

Parastacids (crayfish)

### **Herbivores and Omnivores (Verts)**



### Fish:

Pterygoplichthys disjunctivus (vermiculated sailfin catfish)



Dorosoma cepedianum (gizzard shad)



Heterandria formosa

(least killifish)



Mugil cephalus (striped mullet)



Erimyzon sucetta (lake chubsucker)



Menidia beryllina (inland silverside)



Pseudemys suwanniensis (suwannee cooter)



Notemigonus crysoleucas (golden shiner)



*Percina* sp. (darter)

### **Turtles:**



Chelydra serpentina (common snapping turtle)

On and a second second

(coastal shiner)



Gambusia affinis (mosquito fish)



Pteronotropis hypselopterus

(sailfin shiner)

Pseudemys nelsoni (Florida redbelly cooter)

Pseudemys peninsularis (peninsular cooter)

### **Secondary Consumers**

### Invertebrates:



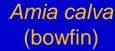
Belostomatidae, Gerridae, Nepidae (predaceous water bugs)



Odonata larvae (dragonfly larvae)



Pisauridae (fishing spiders)





Aphredoderus sayanus

Fish:

Lucania goodei (bluefin killifish)



Ameiurus spp.



(catfish) Micropterus salmoides (Florida largemouth bass) Lepomis spp.



(sunfish)

(pirate perch) **Turtles/Snakes:** 



Sternotherus minor (loggerhead musk turtle)



S. Odoratus (common musk turtle)



Agkistrodon piscivorus (cottonmouth)



Nerodia fasciata (banded watersnake)



N. taxispilota (brown watersnake)

### **Top predators**

Fish:



Micropterus salmoides (Florida largemouth bass)





Esox niger (chain pickerel)



Lepisosteus platyrhincus (Florida gar)



Lepisosteus osseus (longnose gar)

### **American alligator:**





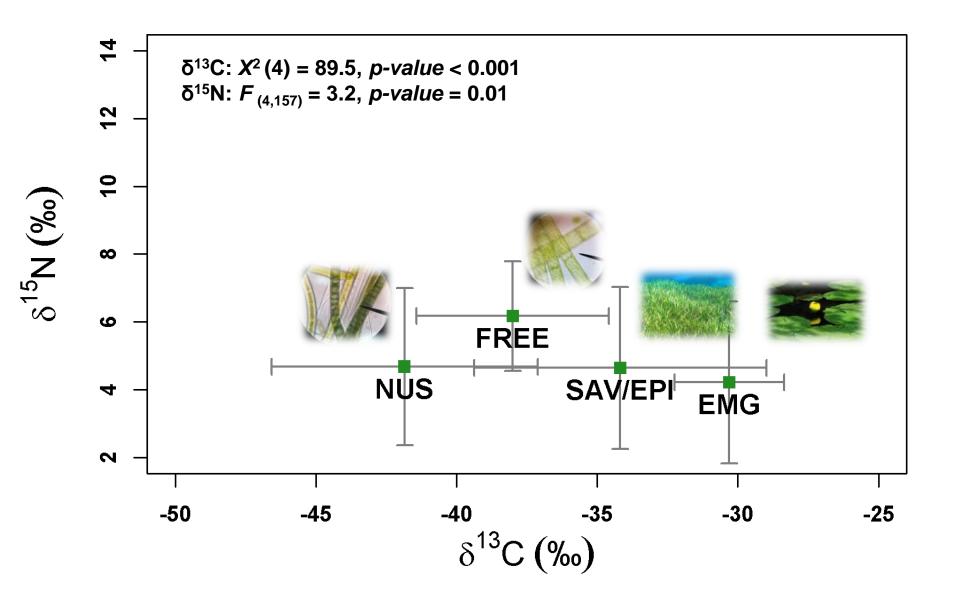




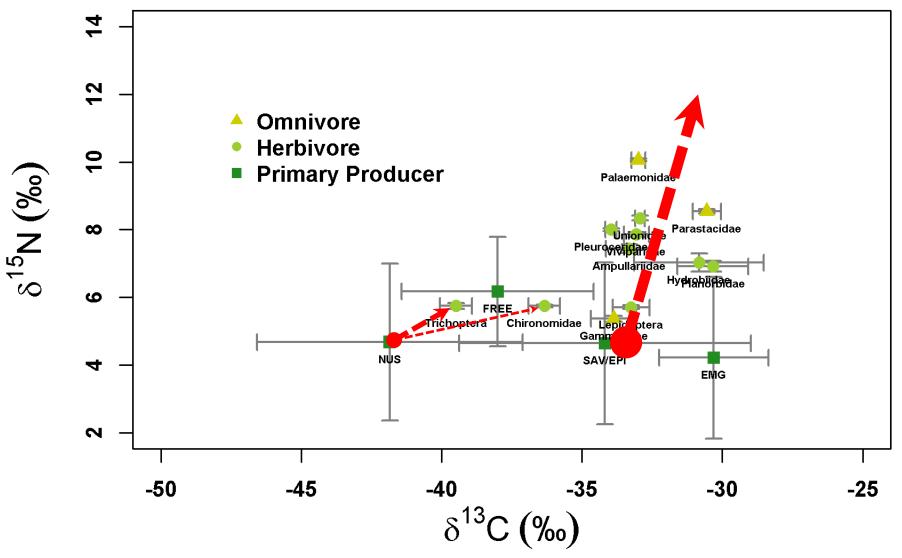


Q1-What are the major pathways of energy flow and material transport in Florida spring ecosystems?

### **Primary Producer Groups (Sources)**

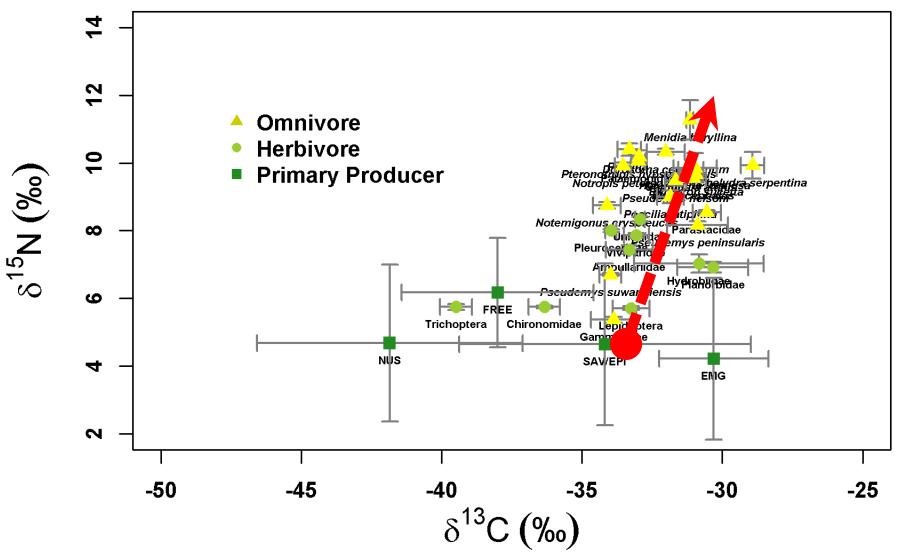


### **Herbivores and Omnivores (inverts)**



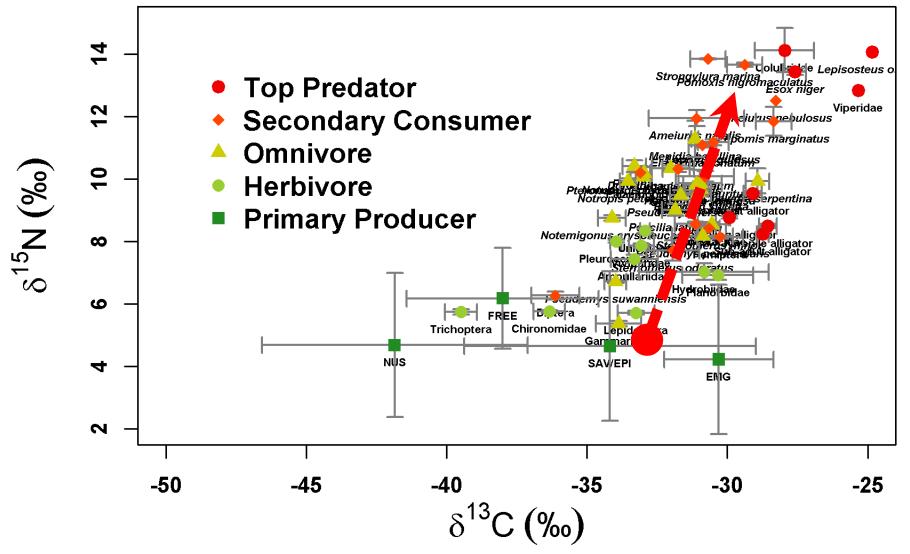
Flow of energy and material transport ----

### **Herbivores and Omnivores**



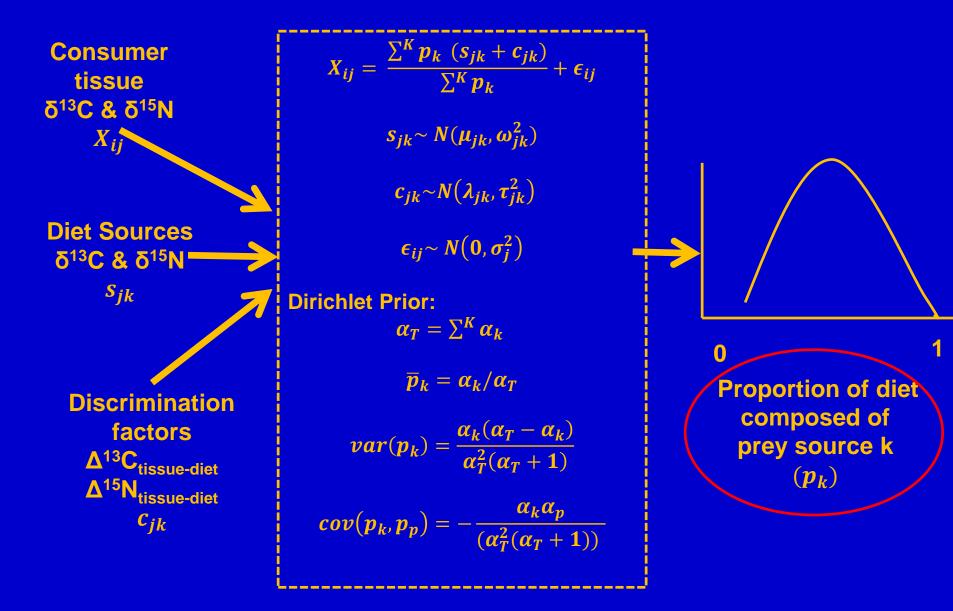
Flow of energy and material transport ----

### **Secondary Consumers and Top Predators**



Flow of energy and material transport ----

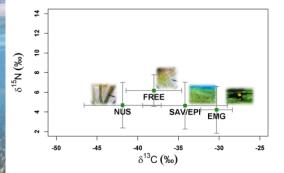
### **Bayesian Isotopic Mixing Model: SIAR**



#### Parnell et al. 2010

# **Mixing Model Results**

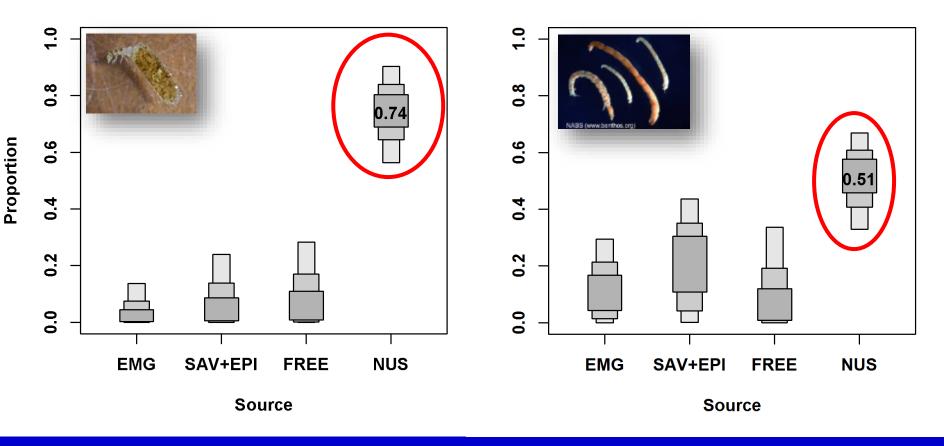
- Four end-member model
  - Sources: NUS, FREE, SAV+EPI, and EMG
  - 28 herbivore and omnivore taxa



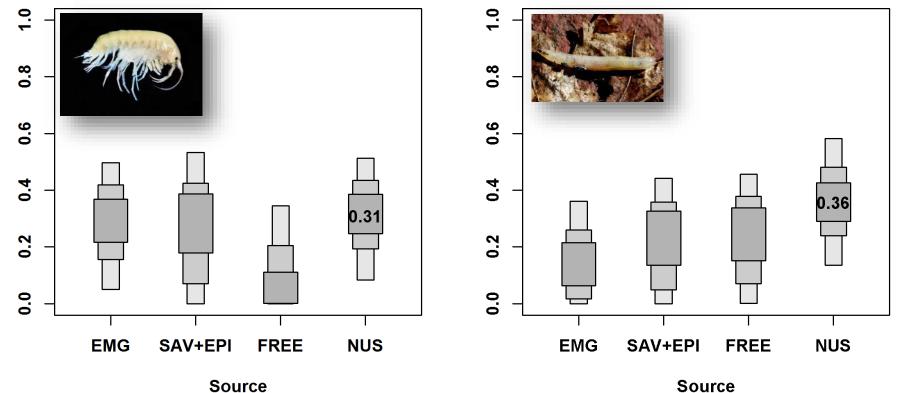
- Contribution of nuisance algae to consumers' diets
  - Median range = 0.04 0.74
    - 8 consumer diets predicted to contain > 30 % nuisance algae

Trichoptera (caddisfly larvae)

Chironomidae (midge larvae)



#### Amphipoda (Gammaridae)

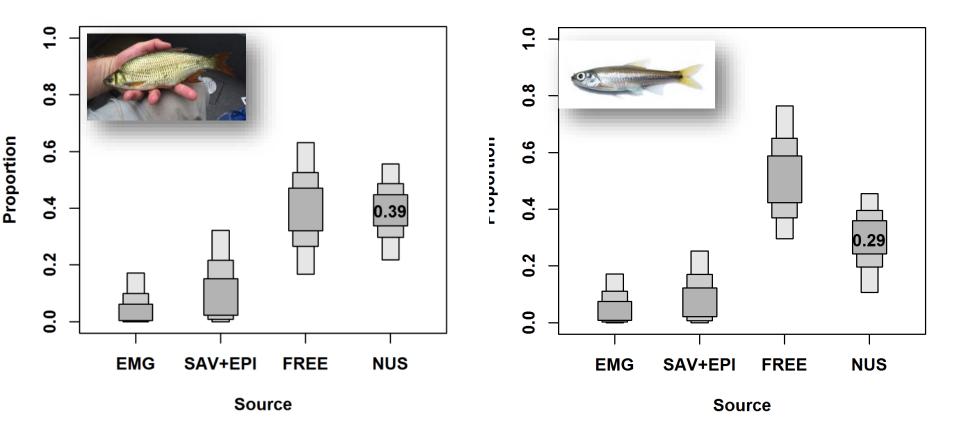


Rhagionidae (snipe fly larvae)

Proportion

Notemigonus crysoleucas (golden shiner)

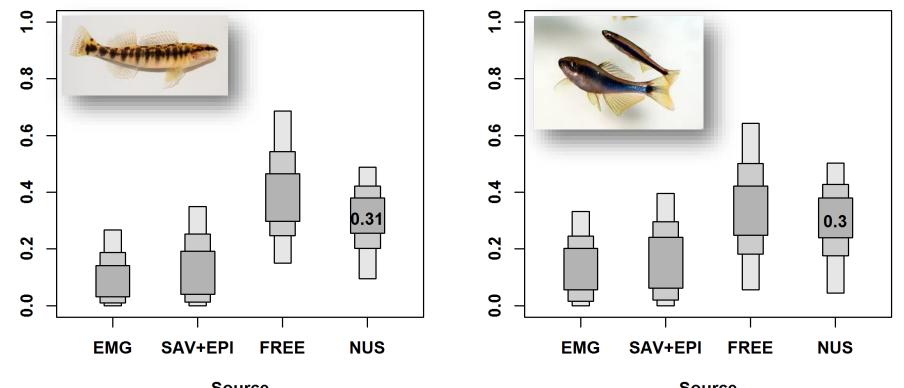
Notropis petersoni (coastal shiner)



Percina sp. (darter)

Proportion





Source

Source

# What can classical dietary data tell us?

Evidence of algal consumption? Who is eating the algal grazers?

# Stomach Content Analysis (SCA)

### **Fish**













**Turtles** 

(scat)



### Alligators

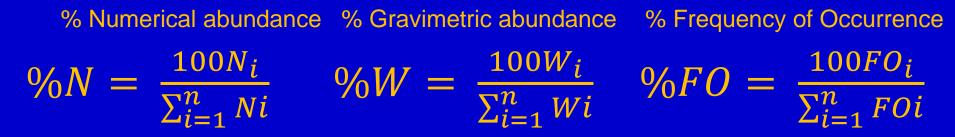




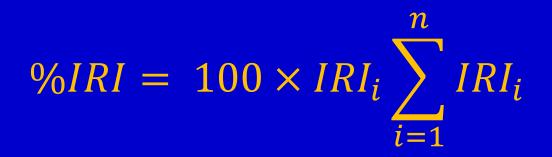


### **Diet Quantification**

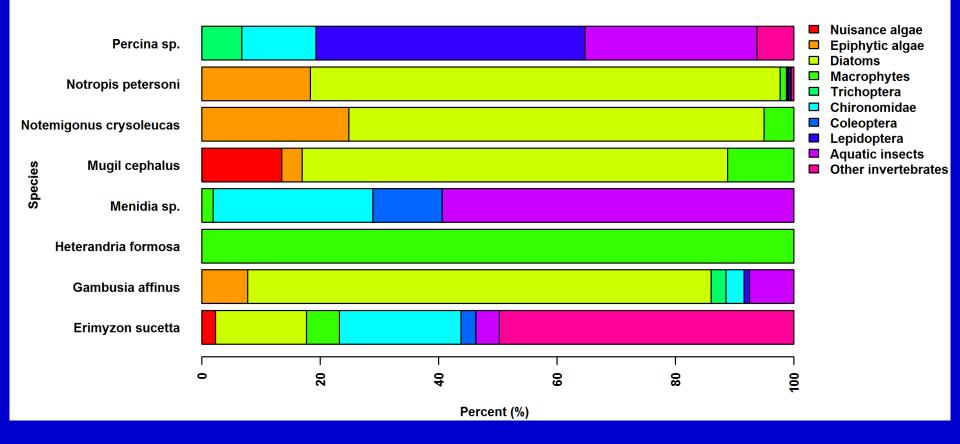
### Percent Index of Relative Importance (%IRI)



 $IRI_{i} = \% FO_{i} (\% W_{i} + \% N_{i})$ 

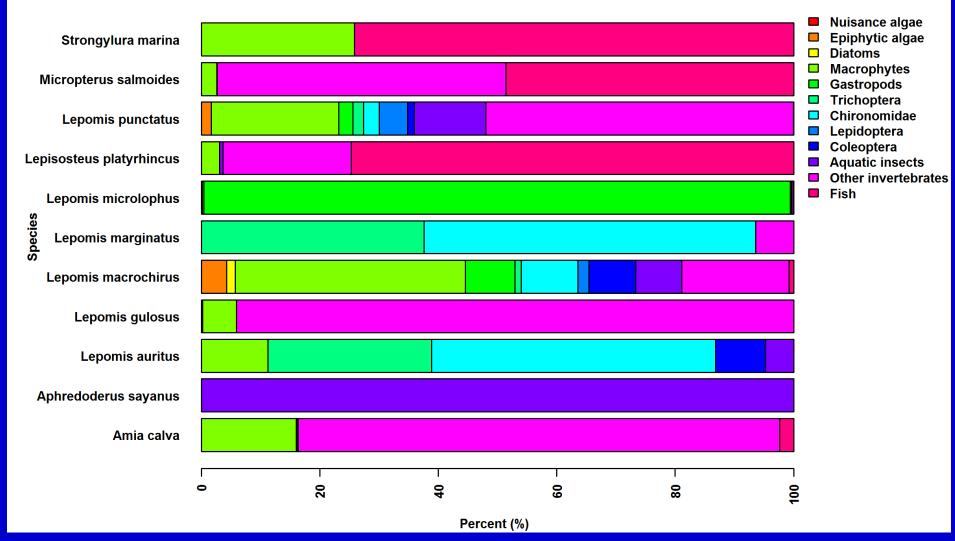


#### % IRI Herbivorous and Omnivorous Fish



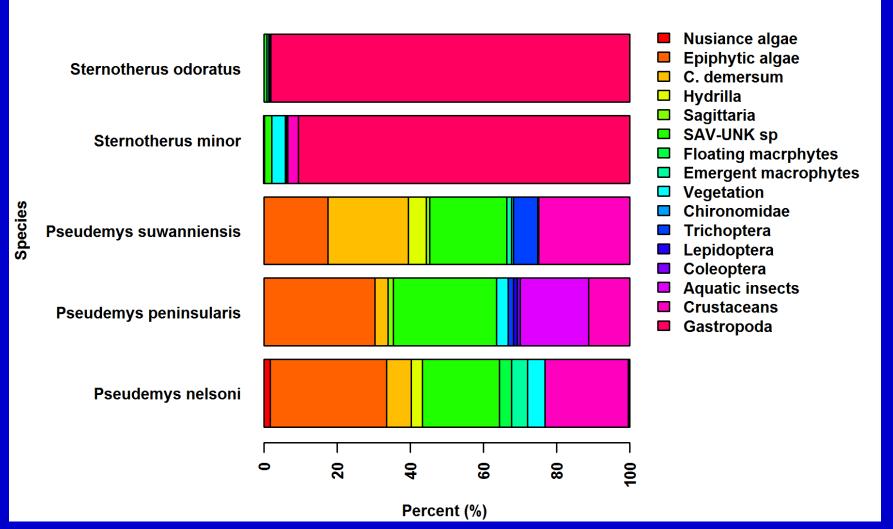
- 1. Diatoms highly important resource.
- 2. Trichopterans and chironomids relatively unimportant prey.
- 3. Little evidence of nuisance algal consumption.

#### % IRI Predatory Fish



- 1. L. marginatus and L. auritus major predators of trichopterans.
- 2. L. microlophus major gastropod predator.
- 3. Other invertebrates (i.e., decapods, amphipods) and fish are primary prey for most species

#### % IRI Turtles



- 1. S. odoratus and S. minor chiefly predators of small benthic gastropods (i.e., physids, hydrobiids, planorbids)
- 2. River cooters (*Pseudemys* spp.) mainly consume macrophytes and to lesser extent small invertebrates.

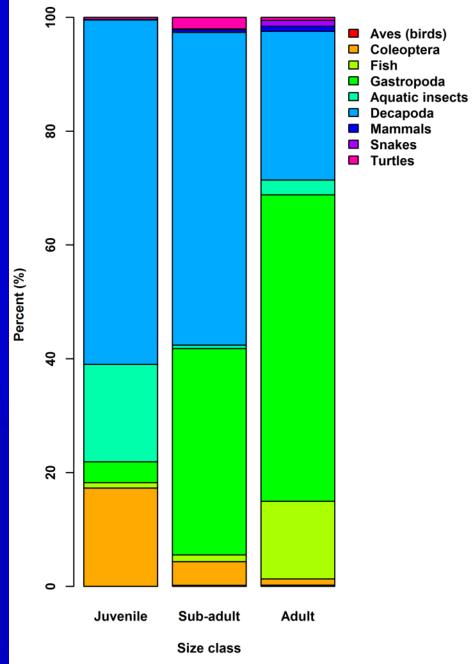
# Alligators







% IRI Alligator mississippiensis



### Conclusions

- Nuisance filamentous contributes little to aquatic food web
  - Few grazers heavily rely of nuisance algae – Inverts: Trichopterans>Chironomids>Rhagionids>Amphipods>Lepi dopterans>Gastropods\*
    - Verts: Shiners > Darters
- Major predators of algal grazers include Redear Sunfish, other Sunfish species, and kinosternid turtles
- Alligators are not 'Apex predators' rather they primarily feed on species occupying lower trophic levels (i.e., gastropods, decapods, insects)

### Determining Grazing Rates of Herbivores on Dominant Macrophyte and Macroalgae Taxa





Planorbella scalaris (rams horn snail) Elimia floridensis (rasp elimia)



Viviparus georgianus (banded mystery snail)



Pomaca paludosa / (Florida apple snail)



Procambarus spiculife (spring crayfish)

Palaemonetes paludosus (Eastern grass shrimp)



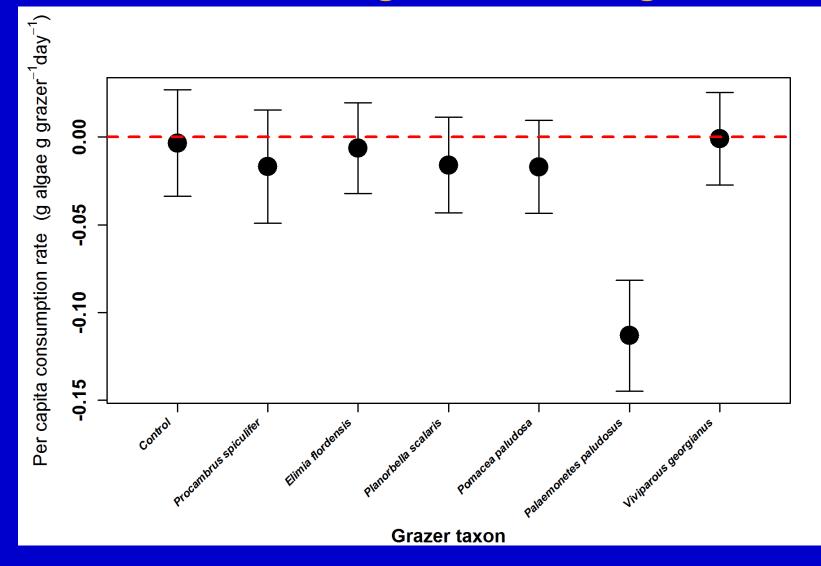


We calculated the per capita consumption rate (g vegetation g grazer<sup>-1</sup> day<sup>-1</sup>) as follows:

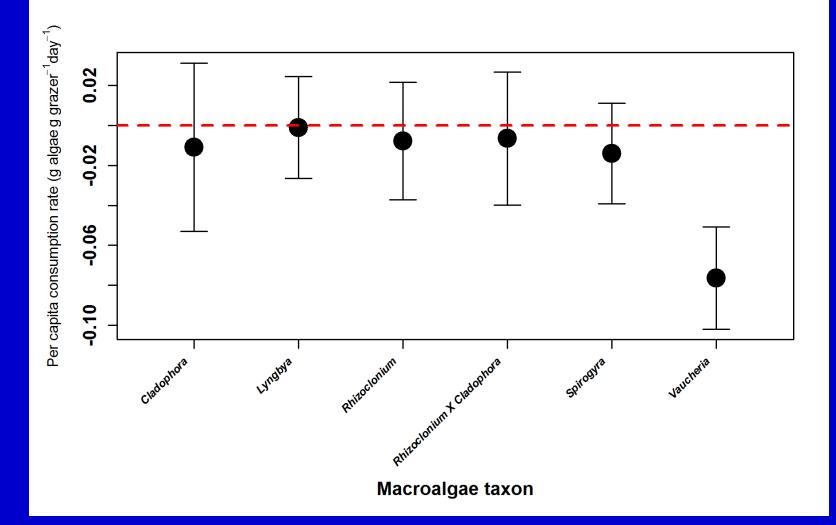
 $Per capita consumption rate = \frac{\frac{final \ veg \ mass - initial \ veg \ mass}{final \ grazer \ mass}}{3 \ days}$ 

### More negative values indicate higher rates of consumption

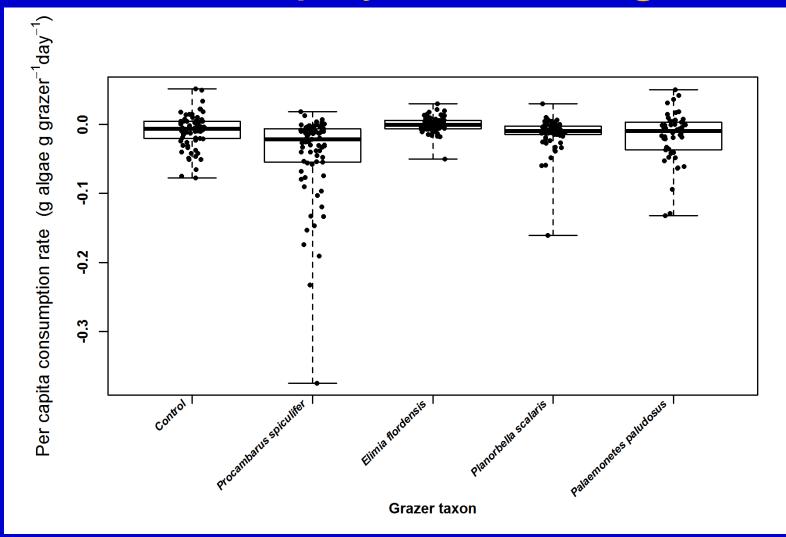
### **Macroalgae Grazing**



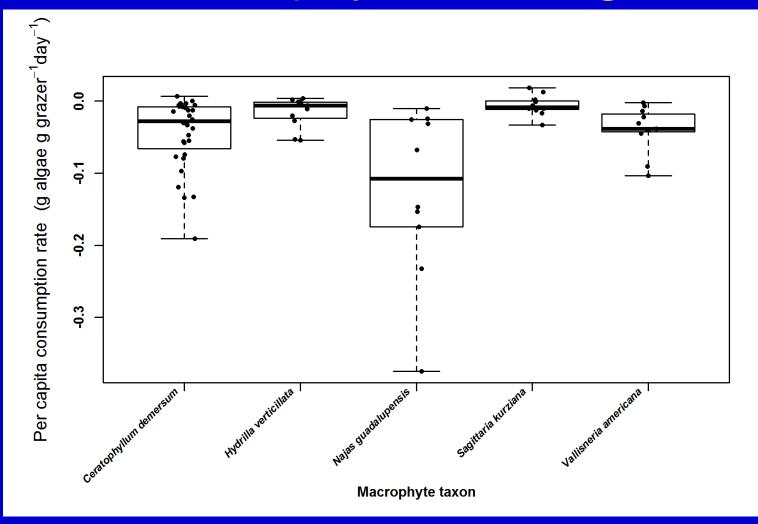
### **Macroalgae Grazing**



# **Macrophyte Grazing**



# **Macrophyte Grazing**

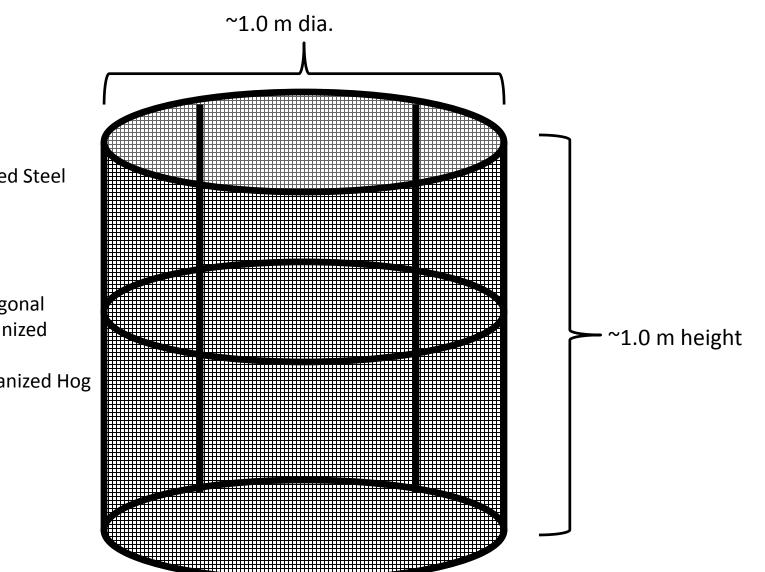


# **Future Directions**

# 1. Complete Grazing Trials

2. Predator Exclusion Experiments

# Cage Design



#### <u>Frame</u>

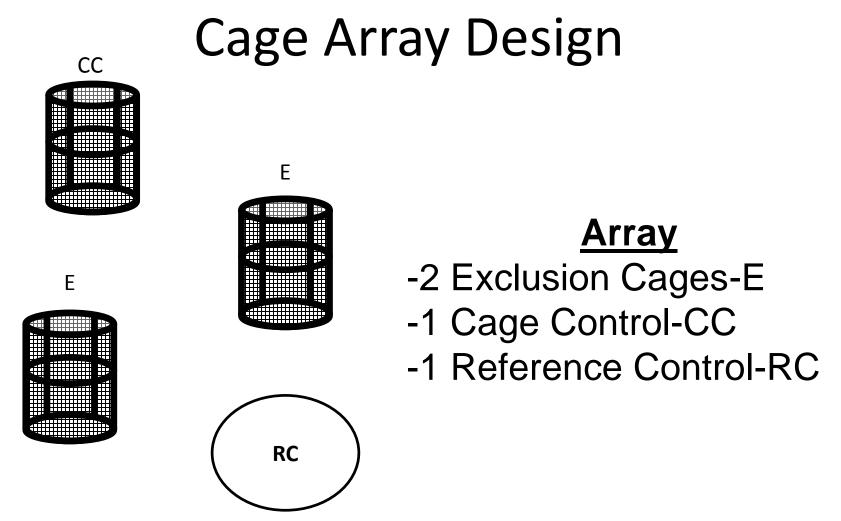
-0.625 cm Galvanized Steel Rods and hoops -All welded

#### <u>Mesh</u>

-2.5 cm mesh Hexagonal -Vinyl Coated Galvanized Steel

-Secured with Galvanized Hog Rings





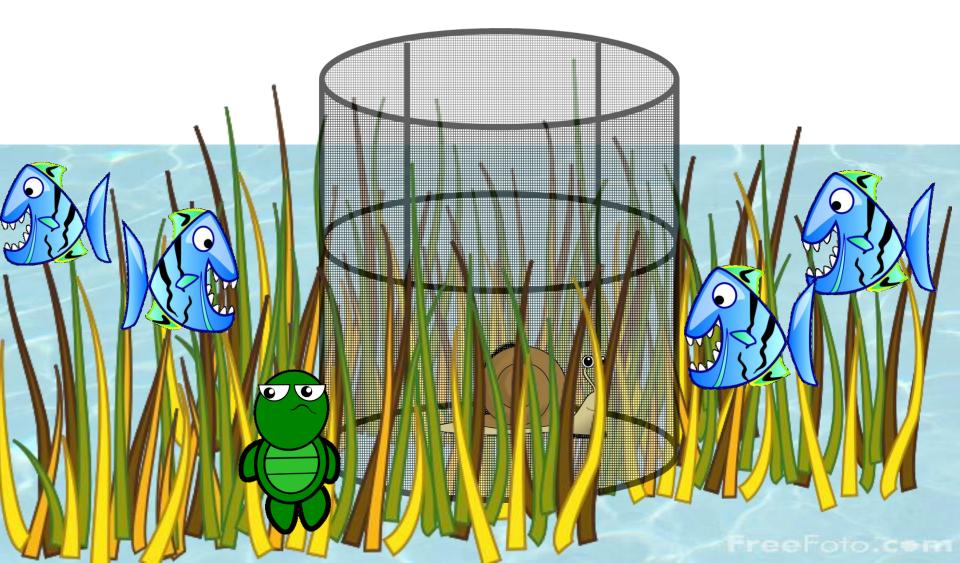
\*Cage Control treatment will have bottom 30 cm of mesh removed from cage to allow organisms uninhibited access while replicating shading and flow effects of true exclusion cages.

\*Reference Control is simply a monitoring area with same footprint as cages.

# Cages may be fully submerged



### Cages may protrude from water's surface



# Acknowledgements

#### **Volunteers and Technicians**

Morgan Farrell Joe Kuehl Jeanelle Brisbane Katherine Ilcken Darvin Johnson Deo Klien Jeff Sowards Cody Godwin Camilo Mojica













### Email: ncboy@ufl.edu