### <u>Annual Report</u>: Springs Ecosystems Supergroup: Hydraulics and Hydrodynamics



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### Integration of Research Elements



### H&HObjectives (from 7/2/14 Meeting)

#### SJRWMD Team

- Predict unsteady water level profiles as f(Q, aquatic vegetation)
- 2. Predict 3D **velocities** in meadowtype (e.g., *Sagittaria*) and canopytype (e.g., *Hydrilla*) SAV
- Develop guidance for selection of Mannings n in different channel types (vegetation type, substrate)
- 4. Develop vegetative resistance and/or turbulence algorithms for
  3-D hydrodynamic modeling

#### <u>UF Team</u>

- 5. Links to 3-D modeling: velocity validation, turbulence msmnts.
- 6. Measure **velocity** and **RTDs** under variety of Q and management
- Quantify the location and magnitude of hyporheic vs. channel storage and exchange
- 8. Identify **critical shear stresses** for entrainment and detachment of filamentous algae

Mostly Data

#### Mostly Models 🔶

### H & H Objectives (simplified)

#### SJRWMD Team

"...determine whether velocity is an important nonnitrate factor influencing the community structure and function of primary producers in the system." (Chapter 6.4)

#### <u>UF Team</u>

#### Mostly Models 🔶

**Mostly Data** 

## H & H Objectives (simplified)



"...determine whether velocity is an important nonnitrate factor influencing the community structure and function of primary producers in the system." (Chapter 6.4)

#### <u>UF Team</u>



**Mostly Data** 

#### Mostly Models 🔶

# Today's Outline

#### 1. Dye trace experiment

- Velocity and residence time distributions
- EFDC model calibration
- 2. Critical velocity/shear stress
  - In-situ flow-ways
  - Optical methods
- 3. EFDC Modeling
  - Domain development
  - Friction/turbulence formulation
  - Initial calibration





# 1. Dye Trace – Background

- <u>**Reach-scale</u>** hydrologic characterization</u>
- Calculate residence time and exchange rates with storage zones

   Nutrient uptake and cycling potential
- Determine if river "behaves" substantially different under different conditions:
  - Channel roughness effects (vegetation build-up downstream?)
  - Water surface profile effects ( $\Delta$  up/downstream river stage?)
  - Couple with EFDC model



# 1. Dye Trace – Background

- Release a tracer; monitor its movement through the system
- Salts, dyes, labeled molecules, etc.
- Measure at set location over time: breakthrough curve (BTC)
- Fitted BTCs → transport properties: advection, dispersion, transient storage...





### 1. Dye Trace – Methods



- Injected 18.9 L of 20% Rhodamine WT at Mammoth Vent
- Tracked dye at 9 fixed stations (3 in-stream fluorometers and 9 ISCO automated samplers)
- Collected 318 grab samples to characterize differential mixing

### Dye Study at Silver Springs March 4, 2015

### 1. Dye Trace – Methods

1-D Advection-Dispersion Equation vs. OTIS

$$\frac{\partial C}{\partial t} = -\frac{Q}{A}\frac{\partial C}{\partial x} + \frac{1}{A}\frac{\partial}{\partial x}(AD\frac{\partial C}{\partial x}) + \frac{q_{LIN}}{A}(C_L - C) + \alpha(C_S - C)$$
$$\frac{dC_s}{dt} = \alpha \frac{A}{A_s}(C - C_s)$$

Q

t

x

α

- main channel cross-sectional area  $[L^2]$
- storage zone cross-sectional area [L<sup>2</sup>]
- main channel solute concentration  $[M/L^3]$
- lateral inflow solute concentration  $[M/L^3]$ 
  - storage zone solute concentration [M/L<sup>3</sup>]
- dispersion coefficient [L<sup>2</sup>/T]

A

 $A_S$ C

 $C_L$ 

 $C_{S}$ 

D

- volumetric flow rate  $[L^3/T]$
- $q_{LIN}$  lateral inflow rate [L<sup>3</sup>/T–L]
  - time [T]
  - distance [L]
  - storage zone exchange coefficient [/T]

# 1. Dye Trace – Methods

- Fitting model to data (get best fit parameters)
- Objective function: e.g., min(SSE), weighting?
- Bayesian parameter fitting
- Parameter identifiability and uniqueness (Kelleher et al. 2013)





Kelleher, C., et al. "Identifiability of transient storage model parameters along a mountain stream." *Water Resources Research* 49.9 (2013): 5290-5306.



Figure: Ed Carter, SJRWMD



- Three <u>upstream</u> peaks (2 back channel flowpaths)
- Full mixing not achieved by 1200 m





- Three <u>upstream</u> peaks (2 back channel flowpaths)
- Full mixing not achieved by 1200 m

- <u>Downstream</u> delay and attenuation
- Comparison to previous study...







- Spring bowl cleared in <6 hours
- Back channel flow: delayed, 2-paths, substantial portion of flow

### 1. Dye Trace – Results: 2009 vs. 2015





### 1. Dye Trace – Results: 2009 vs. 2015







Parameter	2009	2015	Units
Q	15.5	20.0	m <sup>3</sup> /s
L	5300	5300	m
А	73.4	80.8	m <sup>2</sup>
A <sub>S</sub>	18.1	17.3	m <sup>2</sup>
D	10.7	5.8	m <sup>2</sup> /s
α	0.00001	0.00005	1/s
τ	418	357	min
u	0.21	0.25	m/s

### 1. Dye Trace – Results: Standard vs. Bayesian



### 1. Dye Trace – Results: Standard vs. Bayesian



D m^2/min

### 1. Dye Trace – Results: EFDC vs. Measured

- EFDC vs. Measured Data
  - Initial simulation of pulsed dye release at Mammoth Vent
  - Compare to data observed from field experiment...





































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### 2. Critical Velocity/Shear Stress



### 2. Critical Velocity/Shear Stress



King, Sean A. "Hydrodynamic control of filamentous macroalgae in a sub-tropical spring-fed river in Florida, USA." Hydrobiologia 734.1 (2014): 27-37.

#### A. Flow-ways

- Submerged structure placed in flat areas with relatively uniform flow
- Deployed with "control" structure for comparison
- Modify opening to channel or exclude flow
- Measure velocity profiles, cover, downstream transport
- <u>Status</u>: scouting locations, developing prototype...



### **B. Optical Methods**

- <u>Goal</u>: collect algal cover and velocity data over wide area to explore correlations and critical velocity
- Current methods impractical for high resolution, spatially distributed data and relies on human estimation
- We seek a rapid, quantitative method to cover large areas (image processing)









$$Color = \log(Band \ 1) - \log(Band \ 2) = \log\left(\frac{Band \ 1}{Band \ 2}\right)$$
$$ColorBG = \log(B) - \log(G) = \log\left(\frac{B}{G}\right)$$
$$ColorGR = \log(G) - \log(R) = \log\left(\frac{G}{R}\right)$$







#### **B2.** Chromaticity – Pixel-based



- E.g., chromaticity of SAV does not overlap with epiphytic algae
- For 50% algal cover, chromaticity concentrated in the same locations
- Adding spatial (x,y) information allows image clustering



Chromaticity pixel distributions of various algal covers

#### **B2.** Chromaticity – Pixel-based





<u>**Itchetucknee</u>**: Three Clusters (SAV, benthic algae, bottom)</u>



<u>**Rainbow**</u>: Two Clusters (SAV, benthic algae)





<u>Silver</u>: Two Clusters (SAV, benthic algae)

#### **B1. Average Image Color Shift**

• Good correlation with both BG and GR





- Good correlation with both BG and GR
- Training image and field data: similar slope





- Good correlation with both BG and GR
- Training image and field data: similar slope
- Both B-G color and algal cover ~ velocity









### 2. Critical Velocity – Next Steps



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# 3. EFDC Modeling – Background

- Contemporary trends: declining discharge and increasing pool elevation
- Flow  $\rightarrow$  Vegetation  $\rightarrow$  Flow
- <u>EFDC</u>: 3-D hydrodyanamic model to incorporate interactions between velocity, discharge, stage and flow resistance (veg)



From Baird and Johnson, 2014.

# 3. EFDC Modeling – Background

Apparent shift in stage-discharge relationship in Silver River:

- 1. Increased spatial coverage of submersed aquatic vegetation?
- 2. Expansion of hydrilla in the lower Silver and Ocklawaha?
- 3. Reconfiguration of vegetation under low discharge?

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Figure by Ed Carter

$$\partial_{t}(mHu) + \partial_{x}(m_{y}Huu) + \partial_{y}(m_{x}Hvu) + \partial_{z}(mwu) - (mf + v\partial_{x}m_{y} - u\partial_{y}m_{x})Hv$$
  
$$= -m_{y}H\partial_{x}(g\zeta + p) - m_{y}(\partial_{x}h - z\partial_{x}H)\partial_{z}p + \partial_{z}(mH^{-1}A_{V}\partial_{z}u) + Q_{u} - c_{t}\sqrt{u^{2} + v^{2}}umH$$
(1)

$$\partial_{t}(mHv)| + \partial_{x}(m_{y}Huv) + \partial_{y}(m_{x}Hvv) + \partial_{z}(mwv) + (mf + v\partial_{x}m_{y} - u\partial_{y}m_{x})Hu$$
  
$$= -m_{x}H\partial_{y}(g\zeta + p) - m_{x}(\partial_{y}h - z\partial_{y}H)\partial_{z}p + \partial_{z}(mH^{-1}A_{V}\partial_{z}v) + Q_{v} - c_{t}\sqrt{u^{2} + v^{2}}vmH$$
(2)

$$\partial_z p = -gH(\rho - \rho_0)\rho_0^{-1} = -gHb$$
 (3)

 $\partial_t(m\zeta) + \partial_x \bigl( m_y H u \bigr) + \partial_y (m_x H v) + \partial_z (mw) = 0$ 

$$\partial_t(m\zeta) + \partial_x\left(m_y H \int_0^1 u dz\right) + \partial_y\left(m_x H \int_0^1 v dz\right) = 0$$

 $\rho = \rho(p, S, T)$ 



#### **Environmental Fluid Dynamics Code**

- 3-D, vertically hydrostatic, free surface, turbulent-averaged flow equations
- Dynamically-coupled transport equations for turbulent kinetic energy, turbulent length scale, salinity and temperature
- Applied to entire Silver River (finer scale) and Lower
   Ocklawaha from Moss Bluff to Eureka (coarser scale)



#### **1. Defining the shoreline**

- 2. Defining bottom type
- 3. Model grid development
- 4. Model formulation







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Bare	Sandy, rocky, or muddy bottom with less than 5% rooted vegetation. Logs may be present.
Patchy	Clumped, thin, or widely spaced vegetation.
Vegetated	Continuously vegetated with the bottom mostly obscured; open water above canopy deeper than 1 m.
Heavily Vegetated	Continuously vegetated with the bottom mostly obscured; vegetation takes up the majority of the water column.
Topped Out	Vegetation reaches completely to the surface; emergent vegetation may be present.
Trees	Extensive roots and trunks of cypress and other trees.

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- Curvilinear, orthogonal grid (developed jointly by Jones Edmunds, Janicki, SJRWMD)
- 13,439 horizontal cells; 8 vertical cells: 107,512 total
- Cell size variable; average horizontal cell length = 5.8 m



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# 3. EFDC Modeling – Results

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### 3. EFDC Modeling – Results

#### Hydrodynamic Model Test: May 2014



- 17 m<sup>3</sup> s<sup>-1</sup> (605 cfs)
- Downstream stage 10.75 m (35.27 ft) at Conner
- Compare use of (unrealistic) Manning's n vs. vegetation algorithms (SAV drag)
- Uncalibrated...

### 3. EFDC Modeling – Results

#### Hydrodynamic Model Test: May 2014



- Sensitivity analysis:
  - Veg. parameters: density and <u>height</u>
  - Discharge
  - Reconfiguration...

# 3. EFDC Modeling – Initial Conclusions

Apparent shift in stage-discharge relationship in Silver River:

- 1. Increased spatial coverage of submersed aquatic vegetation?
- 2. Expansion of hydrilla in the lower Silver and Ocklawaha?
- 3. Reconfiguration of vegetation under low discharge?

Whitford, 1952:

"After the first mile Silver Springs run becomes narrow and the banks heavily wooded. It also receives some brown water down run. Consequently *about 2 <sup>1</sup>/<sub>2</sub> miles from the boil flowering plants largely disappear* probably due to reduced light. Mats of Vaucheria with some filamentous blue-green algae, and a few of the usually dominant diatoms, are abundant in the shallows. The deeper channel has *relatively little plant life*."

Odum,1957:

"Except for its thick bed of rich muck Silver River would be a rushing canal through a pipe of limestone rock. *Further downstream below the study area it is of this nature*"

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...estimated velocity = 0.21 m/s during Odum study mud, mud/sand, sand, rock: 0.08, 0.11, 0.16 and 0.22 m/s & "little or no SAV" >0.25 m/s (Hoyer et al. 2004)

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- 1. Increased spatial coverage of submersed aquatic vegetation?
- 2. Expansion of hydrilla in the lower Silver and Ocklawaha?
- 3. Reconfiguration of vegetation under low discharge?
- Velocity may play a role in determining vegetative structure and density, especially downstream (e.g., Odum)
- Evidence for recent expansion of vegetation cover in Silver River is not convincing (Duarte et al. 1990; FWC 2014)
- Hydrilla in Ocklawaha? Only recently observed (2011; FWC 2014) and seasonally removed via discharge?
- Vegetation reconfiguration? Model sensitivity to veg. height; lower discahrge/velocity after prolonged drought (1999-2000)
- Likely a combination of all three...

# Thanks! Questions?

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